

# 2×Taq MasterMix

**Cat. No. :** CW0716S (1 mL)  
CW0716M (5 mL)

**Shipping and Storage :** Stored at -20°C.

## Components

Component	CW0716S	CW0716M
	1 mL	5 mL
2×Taq MasterMix	1 mL	5×1 mL
ddH <sub>2</sub> O	1 mL	5×1 mL

Note: 2×Taq MasterMix contains Taq DNA polymerase, 3 mM MgCl<sub>2</sub>, and 400 μM dNTP mix.

## Quality Control

It has no exogenous nuclease activity after inspection; no residual host DNA was detected by PCR method; it can effectively amplify single-copy genes in a variety of genomes.

## Introduction

2×Taq MasterMix is a convenient premixed 2×concentrated solution for PCR which include Taq DNA Polymerase, PCR Buffer, Mg<sup>2+</sup>, dNTPs, PCR Stabilizer and PCR Intensifier. As the unique MasterMix makes the whole reaction system quite stable, more than 98% PCR amplification and complicated templates can be performed. It is easy to use and can maximatily avoid the personal error and contamination. As dye is not contained in the mixture, it is recommended to add loading buffer to the PCR product for electrophoresis. Taq MasterMix catalyzes the non-template directed addition of an adenine residue to the 3'-end of both strands of DNA molecules to make it suitable for TA cloning.

## Precautions

The following basic protocol serves as a general guideline and a starting point for any PCR amplification. Optimal reaction conditions (incubation times and temperatures, concentration of Taq MasterMix, primers, and template DNA) vary and need to be optimized according to the template, primer structure and target fragment size.

1. Prepare the reaction mix to 50  $\mu$ L according to the following table.

Reagent	50 $\mu$ L PCR reaction	Final Concentration
2 $\times$ Taq MasterMix	25 $\mu$ L	1 $\times$
Forward Primer, 10 $\mu$ M	2 $\mu$ L	0.4 $\mu$ M
Reverse Primer, 10 $\mu$ M	2 $\mu$ L	0.4 $\mu$ M
Template DNA	<0.5 $\mu$ g	<0.5 $\mu$ g/reaction
ddH <sub>2</sub> O	Up to 50 $\mu$ L	

**Note:** The recommended primer concentration for PCR is between 0.1-1.0  $\mu$ M per primer. The use of higher concentrations of primers can have for the higher amplification effect. A low primer concentration generally ensures a cleaner product and lower background.

2. PCR reaction conditions:

Procedure	Temperature	Time
Pre-denaturation	94°C	2 min
Denaturation	94°C	30 s
Annealing	55-65°C	30 s
Extension	72°C	30 s
Final extension	72°C	2 min

} 25-35 cycles

**Note:**

- 1) The recommended annealing temperature is about 5°C below T<sub>m</sub>. If extra bands are observed, higher annealing temperatures should be considered. The absence of product can indicate the need for a lower annealing temperature.
- 2) PCR extension time depends on the target gene sequence and CWBIO Taq DNA polymerase is approximately 1 kb DNA/30 second.
- 3) The number of PCR cycles will basically depend on the downstream application of the PCR product.

This product is for scientific research only, which shall not be used for clinical diagnosis or other purposes.